

Benchmarking deep learning-based methods for RNA 3D structure prediction

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Computational Approaches to RNA Structure and Function

Benasque Science Center, Jul 21 - Aug 03, 2024

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Outline

- Benchmarking **deep learning-based tools** for RNA 3D structure prediction
 - overview of the tools
 - datasets
 - results
- Next steps towards our structure prediction model: our RNA language model - RiNALMo

RNA 3D structure prediction

>8UPT_A Primary structure / sequence GGGGGACGGCGACCAGCGGGUCUCUAAAACCUAGCCAGCGGGGUUCGACGCCCCGGUCUCUCGCCA



• DRfold

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- DRfold
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- DeepFoldRNA
- RhoFold

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Tao Shen, Zhihang Hu, Zhangzhi Peng, Jiayang Chen, Peng Xiong, Liang Hong, Liangzhen Zheng, Yixuan Wang, Irwin King, Sheng Wang, Siqi Sun, Yu Li

- DRfold
- DeepFoldRNA
- RhoFold
- RoseTTAFoldNA

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- DRfold
- DeepFoldRNA
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- DRfold
- DeepFoldRNA
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Nature 630, 493–500 (2024) Cite this article

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Research questions

- which of the tools performs best across different datasets and evaluation metrics?
- do certain design choices and methodologies impact accuracy?
- how well these tools generalize to RNA sequences different from those used in their training?
- can we choose the best predicted structure using ARES or Rosetta score?
- (with AF3) how much does having context (other chains from the complex) help in structure prediction?

AlphaFold2's architecture



*Figure adapted from [Jumper et al., 2021]

Data preprocessing



*Figure adapted from [Jumper et al., 2021]

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Data preprocessing: multiple sequence alignment (MSA)



*Figure adapted from [Jumper et al., 2021]

Data preprocessing: secondary structures (SS)



*Figure adapted from [Jumper et al., 2021]

15

End-to-end vs. predicting geometric restraints



*Figure adapted from [Jumper et al., 2021]

End-to-end vs. predicting geometric restraints

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*Figure adapted from [Jumper et al., 2021]

17

From geometric restraints to structural model



*Figure adapted from [Pearce et al., 2022]

Similarities and differences between RNA models

Tool	end-to-end	uses MSA	uses SS	uses LM
DRfold		×		×
DeepFoldRNA	×			×
RhoFold			×	
RoseTTAFoldNA			×	×
trRosettaRNA	×			×

AlphaFold 3 - main differences

- not only for proteins, AF3 works with RNA chains, ligands and combinations of all three
- changed structure module diffusion module
- working on atom level instead of residue level



Datasets

- Dataset 1 = RNA Puzzles
 - 37 RNAs puzzles 35 and 36 removed since they are in CASP15 dataset
- Dataset 2 = CASP15
 - 12 RNAs 8 natural & 4 synthetic
- Dataset 3 = curated dataset from PDB
 - 190 RNAs
 - published after April 2022 in Protein Data Bank (PDB)
 - clustered with sequence identity 90% only considering ones which are not similar to those prior to April 2022
 - filtering: length < 10nt, resolution > 9Å, %defined residues < 90%, sequences containing only 'N' or 'X'
 - 329 clusters => only 190 without errors for all tools





Results



Results - RMSD



Note:

different datasets = different tool with the lowest RMSD

Results - TM-score



Note:

for Dataset 1 RoseTTAFoldNA is the best (has the highest TM-score), not trRosettaRNA (as in the case of RMSD)

Percentage of RNAs for which each tool was the best

according to RMSD: according to TM-score: AlphaFold3 DeepFoldRNA DeepFoldRNA DRfold 21.6% 29.5% 22.1% 23.2% 4.2% 21.1% RhoFold 8.4% RoseTTAFoldNA AlphaFold3 14.2% 17.4% 8.9% 16.8% 12.6% trRosettaRNA RoseTTAFoldNA trRosettaRNA DRfold RhoFold

Scoring functions - ARES and Rosetta scores

- ARES is a deep learning method for scoring RNA structures (input = PDB file, output = score for given structure lower values mean better structure)
- Rosetta score = rna_score from Rosetta toolkit

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Geometric deep learning of RNA structure									
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Dataset 1 -**RNA Puzzles**

Note:

Rosetta score was the lowest for trRosettaRNA's model for all RNAs





Dataset 2 -CASP15

Note: ARES selects AF3 models as best for all RNA targets.



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Dataset 2 - CASP15





Dataset 3

Note:

again, Rosetta score was the lowest for trRosettaRNA's model for all RNAs



Dataset 3: Interaction Network Fidelity (INF)





Dataset 3: Local Distance Difference Test (IDDT)



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Dataset 3: Clash score



Dataset 3: Clash score



Performance (RMSD) per length group (Dataset 3)



Performance (TM-score) per length group (Dataset 3)



Execution time (for all three datasets)



Examples





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Example: 7WM4_B



Note - colors:

native = bright green DRfold = blue DeepFoldRNA = orange RhoFold = dark green RF2NA = red trRosettaRNA = purple AF3 = cyan

RNA chain as a single chain vs. as part of the complex

- RoseTTAFoldNA has an option of providing protein sequences in which it still predicts only RNA chain(s), but in couple of examples where we tried this the resulting predictions were almost the same
- RoseTTAFoldNA tried out on 10 complexes better only in 50% of cases, not by much



RNA chain as a single chain vs. as part of the complex

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- RoseTTAFoldNA tried out on 10 complexes better only in 50% of cases, not by much
- AlphaFold3 for these 10 complexes better for 9/11 chains, for 7YCH_B 91% lower RMSD in complex prediction
- Not sure how trustworthy AlphaFold3 results are (it could be trained on these examples)



Conclusion

- Q: Which of the tools performs best across different datasets and evaluation metrics? A: No unique tool, depends on the use case.
- Q: Do certain design choices and methodologies impact accuracy? A: Unable to answer, no direct connections.
- Q: How well these tools generalize to RNA sequences different from those used in their training?
 A: The best tool on generalization dataset (Dataset 3) is DeepFoldRNA across most metrics.
- Q: Can we choose the best predicted structure using ARES or Rosetta score?
 A: According to our tests, no.
- Q: How much does having context help in structure prediction? A: In case of AlphaFold3, currently it seems a lot, but in case of RoseTTAFoldNA, not that much.

RiNALMo: RNA language model

Pafaol Josip



Rafael Josip Penić

Tin Vlašić

• 36M unannotated ncRNA sequences, 650M parameters

BERT-style language model pretrained using masked language modeling

Motivation: success of protein language models



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Secondary structure prediction

- Finetuned RiNALMo embeddings + small CNN prediction head
- RiNALMo helps to generalize well on RNA families not seen in the training dataset unlike other deep learning methods
 *Dataset obtained from [Szikszai et al., 2022]

Test Family	RNAstructure	CONTRAfold	RiNALMo	RNA-FM	MXfold2	UFold
5S rRNA	0.63	0.67	0.88	0.52	0.54	0.53
SRP RNA	0.63	0.60	0.70	0.25	0.50	0.26
tRNA	0.70	0.76	0.93	0.78	0.64	0.26
tmRNA	0.43	0.44	0.80	0.29	0.46	0.40
RNase P RNA	0.55	0.60	0.80	0.30	0.51	0.41
Group I intron	0.54	0.59	0.66	0.16	0.45	0.45
16S rRNA	0.57	0.60	0.74	0.13	0.55	0.41
Telomerase RNA	0.50	0.54	0.12	0.08	0.34	0.80
23S rRNA	0.73	0.75	0.85	0.17	0.64	0.45
Mean	0.59	0.62	0.72	0.30	0.51	0.44

Future directions

- Currently pretraining a 1.6B parameter RiNALMo on ~100M RNA sequences
- Multimodal pretraining including chemical probing data
- 3D structure prediction model leveraging RiNALMo sequence embeddings

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Collaborators:





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Mile SIKIC, Group Leader

Šikić Lab



https://sikic-lab.github.io

THANK YOU

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