A-to-I RNA editing in *D. melanogaster* alternative RNA structure expression and its role in alternative splicing

Irmtraud M. Meyer

#### MDC-BIMSB & Freie Universität, Berlin, Germany

Benasque, 26. July 2018





## A-to-I RNA editing in *D. melanogaster*:

- abundant in mammals, but also fruit fly
- carried out by ADAR proteins (ADARs)
- requires double-stranded RNA (dsRNA)
- seemingly done in a sequence-unspecific way
- one dsRNA can have multiple editing sites
- ! different copies of same transcript in the same cell can be edited differently
- ! editing can turn dsRNA into ssRNA

**Hypothesis:** Can alternative splicing be regulated by A-to-I editing via RNA structure changes ?



# Result #1: analysis pipeline for detecting A-to-I editing sites

Key features:

- (1) filter against artifacts due to mapping and sequencing errors
- (2) explicitly capture ADAR-specific features such as requirement for dsRNA and that editing sites tend to occur in clusters
- (3) leverage the statistical power derived from the size and number of our input data sets
- (4) devise probabilistic pipeline whose parameters can be trained and whose predictions can be ranked

# Result #1: analysis pipeline for detecting A-to-I editing sites



◆□▶ ◆□▶ ◆ □▶ ◆ □▶ ○ □ ○ ○ ○ ○

- data: stranded, paired-end RNA-seq data (29 libraries, 10 tissue types)
- results:
- pprox 2000 new editing sites
- A-to-I editing especially in head and CNS
- majority of editing cites in non-coding regions
- editing sites mostly tissue-specific
- alternatively spliced genes higher chance of being edited (p-value  $< 2 \cdot 10^{-15}$ )
- editing 3 x more likely in altern. spliced exons (p-value  $< 2 \cdot 10^{-15}$ )



- data: stranded, paired-end RNA-seq data (29 libraries, 10 tissue types)
- results:
- pprox 2000 new editing sites
- A-to-I editing especially in head and CNS
- majority of editing cites in non-coding regions
- editing sites mostly tissue-specific
- alternatively spliced genes higher chance of being edited (p-value  $< 2 \cdot 10^{-15}$ )
- editing 3 x more likely in altern. spliced exons (p-value  $< 2 \cdot 10^{-15}$ )



▲ロト ▲帰ト ▲ヨト ▲ヨト 三日 - の々ぐ

- data: stranded, paired-end RNA-seq data (29 libraries, 10 tissue types)
- results:
- pprox 2000 new editing sites
- A-to-I editing especially in head and CNS
- majority of editing cites in non-coding regions
- editing sites mostly tissue-specific
- alternatively spliced genes higher chance of being edited (p-value  $< 2 \cdot 10^{-15}$ )
- editing 3 x more likely in altern. spliced exons (p-value < 2 · 10<sup>-15</sup>)



< □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > < □ > <

- data: stranded, paired-end RNA-seq data (29 libraries, 10 tissue types)
- results:
- pprox 2000 new editing sites
- A-to-I editing especially in head and CNS
- majority of editing cites in non-coding regions
- editing sites mostly tissue-specific
- alternatively spliced genes higher chance of being edited (p-value  $< 2 \cdot 10^{-15}$ )
- editing 3 x more likely in altern. spliced exons (p-value < 2 · 10<sup>-15</sup>)



- data: stranded, paired-end RNA-seq data (29 libraries, 10 tissue types)
- results:
- pprox 2000 new editing sites
- A-to-I editing especially in head and CNS
- majority of editing cites in non-coding regions
- editing sites mostly tissue-specific
- alternatively spliced genes higher chance of being edited (p-value  $< 2 \cdot 10^{-15}$ )
- editing 3 x more likely in altern. spliced exons (p-value < 2 · 10<sup>-15</sup>)



(日)、

### Result # 3: link between splicing and A-to-I editing



- identify 244 edited regions: editing and change in splicing
- 96 of 244 regions contain evolutionarily conserved RNA structure

イロト 不得 トイヨト イヨト

-

### Result # 3: link between splicing and A-to-I editing



- identify 244 edited regions: editing and change in splicing
- 96 of 244 regions contain evolutionarily conserved RNA structure

▲ロト ▲帰ト ▲ヨト ▲ヨト 三日 - の々ぐ

#### Result # 4: proposed model of action:

#### tissue-specific RNA editing of dsRNAs near splice sites

⇒ induced changes in local RNA structure
⇒ induced changes in splicing pattern

[Brugiolo et al., F1000Prime Rep. (2013) 5:9; Mazloomian & Meyer, RNA Biology (2015) 12(12):1391-401; Herzel et al., Nat. Rev. Mol. Cell Biol. (2017) 18(10):637-650; Meyer, Methods (2017) 120:3-16.]

▲ロト ▲帰ト ▲ヨト ▲ヨト 三日 - の々ぐ

#### Result # 4: proposed model of action:

# tissue-specific RNA editing of dsRNAs near splice sites $\Rightarrow$ induced changes in local RNA structure

 $\Rightarrow$  induced changes in splicing pattern

[Brugiolo *et al.*, F1000Prime Rep. (2013) 5:9; Mazloomian & Meyer, RNA Biology (2015) 12(12):1391-401; Herzel *et al.*, Nat. Rev. Mol. Cell Biol. (2017) 18(10):637-650; Meyer, Methods (2017) 120:3-16.]

#### Result # 4: proposed model of action:



tissue-specific RNA editing of dsRNAs near splice sites  $\Rightarrow$  induced changes in local RNA structure  $\Rightarrow$  induced changes in splicing pattern

[Brugiolo et al., F1000Prime Rep. (2013) 5:9; Mazloomian & Meyer, RNA Biology (2015) 12(12):1391-401; Herzel et al., Nat. Rev. Mol. Cell Biol. (2017) 18(10):637-650; Meyer, Methods (2017) 120:3-16.]

◆□▶ ◆圖▶ ★ 圖▶ ★ 圖▶ / 圖 / のへで

# Summary & Acknowledgements:

Key points:

- local RNA structure can be involved in regulating splicing via RNA structure changes Concept of alternative RNA structure expression.
- repeats may play a functional role in regulating splicing

N.B.: 10% of human genome are Alu-repeats. These could constitute potential A-to-I editing sites.



A. Mazloomian

Contact: irmtraud.meyer@mdc-berlin.de



Fondation canadienne nour l'innovatio



ULAR MEDICINE THE HELMHOLTZ ASSOCIATION

MAX DEL BRÜCK CENTER

FOR MOLEC





