SHAPE as a way to evaluate our ability to predict the effects of SNPs on RNA

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*In our CAFA analysis, peaks are proportional to the amount of accessibility-dependent labeling in folded RNA at different positions

*Peaks here represent unfiltered fluorescence intensities of the three variants of FTL 5'UTR that we will focus on today (WT,A196G and U22G)

*The area that is being focused on here is representative of the IRE in the secondary structure of 5'UTR

*We see that even without filtration of datapoints, not only do WT and A196G capillary traces closely follow one another, but also U22G deviates significantly from the WT patterns of peaks shown



SNRNASM (Single Nucleotide Resolution Nucleic Acid Structure Mapping)



Listed below are some example data sets shared by different labs in the ISATab format. You can also obtain a blank ISATab file here and find the ontological references from the tutorial here.

Evaluation of the information content of RNA structure mapping data for secondary structure prediction. DMS and T1 structure probing of the Twort and Tetrahymena group I introns under low and high salt conditions. Atomic accuracy in predicting and designing noncenonical ma structure. DMS probing of a synthetic RNA and three mutants under multiple Mg solution conditions. RNA: Folding and Millisecond Intervals by Synchrotron Hydroxyl Radical Fostprinting. Time Resolved Hydroxyl Radical Fostprinting of the Tetrahymena group I intron when folding in 10mM Mg.

Nonhiererchical Ribonucleoprotein Assembly Supports a Strain-Propagation Model for Protein-Facilitated RNA Folding. SHAPE probing data on maturase bound RNA (Mrs-1-bound RNA) in the presence and absence of Maturase.



SNRNASM is a link farm to Google Spreadsheets

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1	SRP.Domain.IV WT	GGCI IACGCAAGI IAAAACAAAI II IACI ICAGGI ICCG	CAACCAAC	DNA	in vitro synthesis	DMS structure mapping assay	
2	SRP-Domain-IV WT	GGCUACGCAAGUAAAACAAAUUACUCAGGUCCG	GAAGGAAC	DNA	in vitro synthesis	DMS structure mapping assay	
3	SRP.Domain IV WT	COCULACIONA A CLARA A CARALLUACIUCACIUCCO	CAACCAAC	DNA	in vitro synthesis	CMC structure mapping assay	-17
4	SPP.Domain-JV WT	OCCURCE AND A AACAAAUUACUCAGUCCO	GAAGGAAC	DNA	in vitro synthesis	DMC structure mapping assay	
5	SRP.Domain.IV WT	COCULACIONA CI LA AACAAALI LACI ICACCI ICCC	CAACCAAC	DNA	in vitro synthesis	PMC structure mapping assay	
6	SRP-Domain-IV WT	GGCUACGCAAGUAAAACAAAUUACUCAGGUCCG	GAAGGAAC	DNA	in vitro synthesis	DMS structure mapping assay	
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Providing open access to the data and meta-data.

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£.,	0.8715	0.6542	0.6584	0.6222	0.6457	0.6079	0.16	0.806	0.8706	0.3093	0.5344	0.1635	0.2873	0.2875	0.0606	0.7531	1
1	0.6955	0.559	0.5986	0.5436	0.5671	0.5447	0.4625	0.7302	0.7161	0.5068	0.7365	0.4865	0.5967	0.5536	0.3926	0.6964	1
	1.8664	1.628	1.6021	1.6027	1.5791	1.5904	1.6923	1.7694	1.7981	1.673	1.5415	1.4349	1,402	1.1963	1.2652	1.3482	
	0.5771	0.5581	0.5442	0.5203	0.5279	0.5415	0.5615	0.6245	0.6806	0.6048	0.5942	0.5366	0.5657	0.4826	0.4761	0.5146	1
	0.2189	0.1901	0.1989	0.1811	0.1928	0.2153	0.228	0.2507	0.2696	0.2461	0.2509	0.2541	0.2602	0.2194	0.2128	0.2219	1
1	1.7793	1.5389	1.5182	1.4979	1.5299	1.6041	1.733	2.0141	2.1061	1.9912	2.1186	2.0432	2.0747	1.9658	1.8557	1.795	
0	1.9935	1.7228	1.7534	1.7279	1.7632	1.8105	1.9047	2.2399	2.3541	2.1165	2.1329	2.059	2.058	1.9091	1.9125	1.8441	-
1	1.6205	1.4371	1.436	1.4261	1.4325	1.465	1.564	1.8443	1.8927	1.697	1.768	1.6721	1.6922	1.6252	1.6876	1.6561	
2	0.348	0.2546	0.2566	0.241	0.2586	0.2543	0.2636	0.3023	0.3736	0.338	0.3167	0.3208	0.3587	0.297	0.2803	0.3007	1
2	0.3793	0.3132	0.3329	0.3028	0.3227	0.3535	0.3648	0.403	0.4447	0.4018	0.3993	0.4083	0.3897	0.3469	0.3429	0.3958	1
1	1.7865	1.5499	1.5654	1.5429	1.5426	1.5849	1.655	1.918	1.9783	1.7541	1.7435	1.6059	1.6062	1.4915	1.5818	1.7448	
2	2.1472	1.9054	1,9588	1.949	1.9902	2.0058	2,1434	2.5652	2.7523	2.4213	2.4460	2.3517	2.3612	2.247	2.274	2.3596	-
	2.3146	2,1586	2.174	2.2254	2.3059	2.3564	2.4721	3.0049	3.2379	2.8967	2.9065	2.8722	2.9058	2.7029	2.7237	2,6805	1
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Open Questions

- What exactly is SHAPE telling us? Notice we only used it comparatively, i.e. does the structure change and how much?
- Is there information in partial SHAPE (or for that matter any probing) reactivities or do we just need to know the most reactive species?
- Why do we correctly predict that there are major structure disrupting SNPs but no one gets the right ones?
- Are hyper-reactive nucleotides even structurally important or just an artifact of ideal acylation geometry?

